



*Supplement of*

## **Climate change increases riverine carbon outgassing while export to the ocean remains uncertain**

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## 1 1 Respiration of litter and soil carbon

2 The respiration of the un-respired litter carbon and the soil carbon has been calculated analogous  
3 to the LPJmL functions with Eqs. (S1) to (S12)

$$LitC_{unresp_{loss}} = LitC_{unresp_t} \times (1 - e^{-(respi_{litter} \times Tresponse_t)}) \quad (S1)$$

$$LitC_{loss} = LitC_t \times (1 - e^{-(respi_{litter} \times Tresponse_t)}) \quad (S2)$$

$$lrLitC_{unresp_t} = LitC_{unresp_t} - LitC_{unresp_{loss}} \quad (S3)$$

$$lrLitC_t = LitC_t - LitC_{loss} \quad (S4)$$

$$LitC_t = lrLitC_t + lrLitC_{unresp_t} \quad (S5)$$

$$lrSoilC_{fast_t} = SoilC_{fast_t} + respipart_{soilfast} (LitC_{unresp_{loss}} + LitC_{unresp_{loss}}) \quad (S6)$$

$$lrSoilC_{slow_t} = SoilC_{slow_t} + respipart_{soilslow} (LitC_{unresp_{loss}} + LitC_{unresp_{loss}}) \quad (S7)$$

$$SoilC_{fast_{loss}} = lrSoilC_{fast_t} \times (1 - e^{-(respi_{soilfast} \times Tresponse_t)}) \quad (S8)$$

$$SoilC_{slow_{loss}} = lrSoilC_{slow_t} \times (1 - e^{-(respi_{soilslow} \times Tresponse_t)}) \quad (S9)$$

$$srSoilC_{fast_t} = lrSoilC_{fast_t} - srSoilC_{fast_{loss}} \quad (S10)$$

$$srSoilC_{slow_t} = lrSoilC_{slow_t} - srSoilC_{slow_{loss}} \quad (S11)$$

$$SoilC_t = srSoilC_{fast_t} + srSoilC_{slow_t} \quad (S12)$$

4 with  $respipart_{soilfast}$  being the fraction of litter that enters the soil organic carbon pool with fast  
5 respiration and  $respipart_{soilslow}$  being the fraction of litter that enters the soil organic carbon pool  
6 with slow respiration.

7

## 8 2 Mobilization

9 The mobilization takes place heterogeneously in the cell. It occurs first closest to the river. The  
 10 cells are therefore divided into fractions, which size depends on the vicinity to the river, with  
 11 section 6 close to the river.

12



13 Figure S1: Depiction of the fraction of each cell section.

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## 15 3 Sensitivity analysis

### 16 3.1 Initial parameter setting and quality

17 The parameterization of the model builds upon an analysis of the scientific literature. The  
 18 parameters used within the model originate from a number of sources and are of differing  
 19 quality. Table S1 lists all parameters and their sources. In addition to the parameter value, it also  
 20 provides the value ranges and a first quality assessment of the parameter values based on the  
 21 methods used in the relevant studies. The quality was weighted medium to low if the  
 22 measurements took place in a slightly other system, for instance in the *Igapó* instead of *Várzea*,  
 23 or are only based on one single observation. The quality and the relevance of single parameters  
 24 for the simulation outputs are further tested in the sensitivity analysis.

25

26 Table S1: Initial parameter setting. List of parameters and parameter quality (high, medium, low).

<i>parameter name</i>	<i>initial value</i>	<i>unit</i>	<i>source</i>	<i>quality</i>
<b>mobilization</b>				
<i>carboncorr</i>	0.65 ± 0.15	month <sup>-1</sup>	(Worbes, 1997)	high
<i>mobil<sub>litc</sub></i>	0.4 ± 0.1	month <sup>-1</sup>	(Irmler, 1982)	medium
<i>mobil<sub>soile</sub></i>	0.008 ± 0.002	month <sup>-1</sup>	(Irmler, 1982)	low
<i>mobil<sub>p</sub></i>	0.5 ± 0.25	-	(McClain and Elsenbeer, 2001; Johnson et al., 2006)	medium
<b>decomposition</b>				
<i>decomp</i>	0.3 ± 0.1	month <sup>-1</sup>	(Furch and Junk, 1997)	high
<i>decompcorr</i>	0.1 ± 0.01	month <sup>-1</sup>	(Furch and Junk, 1997)	high
<b>respiration</b>				
<i>respi</i>	0.045 ± 0.01	day <sup>-1</sup>	(Cole et al., 2000)	high
<b>outgassing</b>				
<i>co2satur</i>	7.25 to 17.0	-	(Richey et al., 2002)	high

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## 28 3.2 Simulations for sensitivity analysis

29 The model RivCM has been run on a  $0.5^\circ \times 0.5^\circ$  spatial resolution for the period 1901-2003. The  
 30 transient runs have been preceded by a 90-years-spinup during which the climate, CO<sub>2</sub> levels and  
 31 carbon input (litter and soil) of 1901-1930 have been repeated to obtain equilibrium for the  
 32 riverine carbon pools. As input to the terrestrial litter and soil carbon pools, LPJmL results  
 33 produced under the CRU TS2.1 climate (Österle et al., 2003; Mitchell and Jones, 2005) has been  
 34 used. The transient LPJmL runs have been preceded by a 1,000-years-spinup during which the  
 35 pre-industrial CO<sub>2</sub> level of 280 ppm and the climate of the years 1901-1930 have been repeated  
 36 to obtain equilibrium for vegetation, carbon and, water pools. For this analysis, simulations have  
 37 been conducted with an initial parameter setting (see Table S1) and a modified parameter setting  
 38 (Table S2).

39

40 Table S2: List of parameters modified for the sensitivity analysis. All parameters have been  
 41 multiplied with the following factors: 0.1; 0.5; 0.9; 1.1; 1.5; 1.9.

<i>parameter name</i>	<i>original value</i>	<i>modified value</i>
<b>mobilization</b>		
<i>mobil<sub>litc</sub></i>	0.4	0.04; 0.2; 0.36; 0.44; 0.6; 0.76
<i>mobil<sub>soilc</sub></i>	0.008	0.0008; 0.004; 0.0072; 0.0088; 0.012; 0.0152
<i>mobil<sub>p</sub></i>	0.5	0.05; 0.25; 0.45; 0.55; 0.75; 0.95
<b>decomposition</b>		
<i>decomp</i>	0.3	0.03; 0.15; 0.27; 0.33; 0.45; 0.57
<b>respiration</b>		
<i>respi</i>	0.045	0.0045; 0.0225; 0.0405; 0.0495; 0.0675; 0.0855

42

43 The sensitivity analysis aims to estimate the effect of changes in the explaining variables on the  
 44 response variables. The results of these simulations have been analysed with a redundancy  
 45 analysis (RDA). This analysis is, comparable to PCA, an ordination technique which identifies  
 46 the most important separator of a given dataset (including all response variables) and also the  
 47 most important initiator (explaining variables) of dataset's variability. The sensitivity analysis led  
 48 to a partly adapted parameter setting (standard parameters). For evaluation, simulations under the  
 49 standard parameter setting (see Table 3 row 'original value') have been conducted. The results of  
 50 these simulations have been compared to several observed values (Table 4).

51

## 52 3.3 Results of sensitivity analysis

53 The aim of a sensitivity analysis is to estimate which output variable (response variable) is most  
 54 sensitive to changing parameters (explaining variables), and which parameter changes cause the  
 55 largest shifts in the output values.

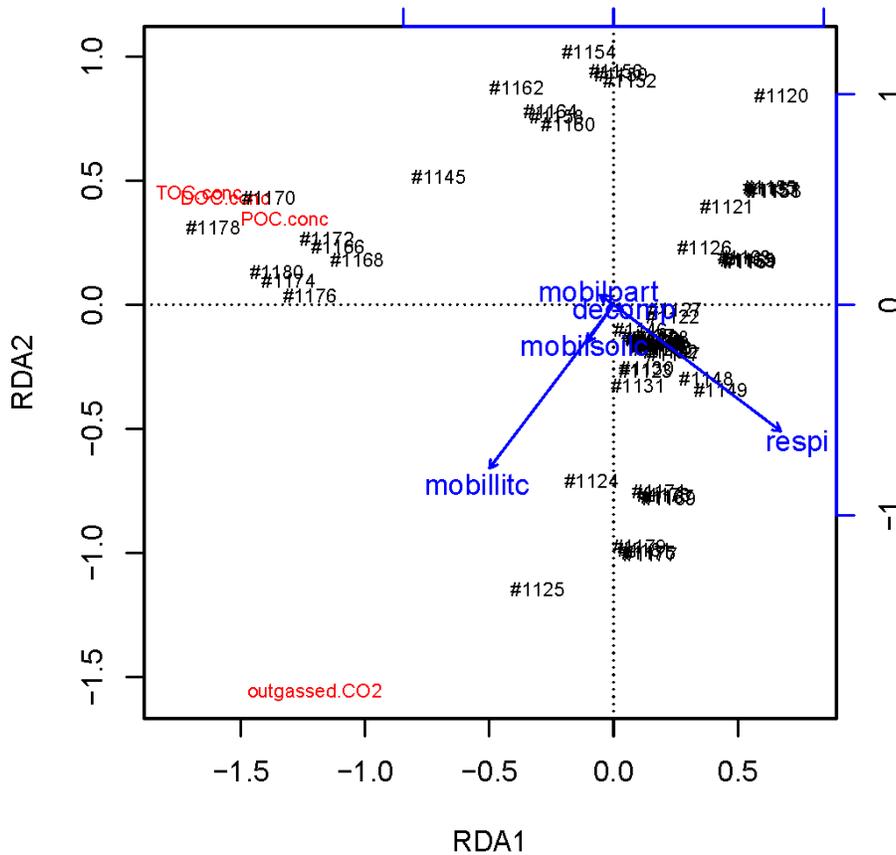
56 To analyse the results of the simulation of the sensitivity analysis a redundancy analysis (RDA)  
57 has been performed. The results of the redundancy analysis (for parameters see Table S2) are  
58 summarized in and Table S3. This analysis shows the effect of the explaining variables, i.e.  
59 parameters, *mobil<sub>litc</sub>*, *mobil<sub>soilc</sub>*, *mobil<sub>p</sub>*, *decomp* and *respi* on the response variables riverine  
60 particulate organic carbon (POC), riverine dissolved organic carbon (DOC), riverine inorganic  
61 carbon (IC) and outgassed carbon. The parameter changes did not cause changes in IC, since it is  
62 only temperature and atmospheric CO<sub>2</sub> dependent. Therefore, IC has not been included in this  
63 analysis.

64 The RDA shows that 79.2% of the variance within the dataset can be described by the explaining  
65 variables (therefore, 20.8% cannot be explained by explaining variables).

66 The first, second, and third axis explain 54%, 18.9%, and 6.3% of the variance within the  
67 dataset, respectively. The variance of the response variables TOC concentration, DOC  
68 concentration, and POC concentration are mainly influenced by the first axis (RDA1) with  
69 loadings (*prop*) of -0.23, -0.21, and -0.18, respectively (depicted in red in Figure S2, listed in  
70 Table S3). This axis is primarily controlled by *respi* (0.79) and *mobil<sub>litc</sub>* (-0.59) (blue arrows in  
71 Figure S2). The variance of the response variable outgassed CO<sub>2</sub> is mainly influenced by RDA2  
72 with a loading of -0.075. This axis is also primarily controlled by *respi* (-0.60) and *mobil<sub>litc</sub>*  
73 (-0.78), but in a swapped order and, in contradiction to RDA1, not in opposite directions. The  
74 third axis (RDA3) mainly influences the response variables POC concentration and DOC  
75 concentration, with loadings of 0.013 and -0.008, respectively. This axis is primarily controlled  
76 by *mobil<sub>p</sub>* (0.81) and *decomp* (-0.59).

77 Therefore, the parameters that explain most of the variance within the dataset are *respi* and  
78 *mobil<sub>litc</sub>*. The parameters *mobil<sub>p</sub>* and *decomp* have only little effect on the variance of the whole  
79 dataset. The most and nearly equally influenced output variables are TOC concentration and  
80 DOC concentration. POC concentration and outgassed CO<sub>2</sub> are only marginally affected.

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Figure S2: Results of the redundancy analysis. Redundancy analysis with all simulations associated with the sensitivity analysis (black numbers). The four output variables (red) have been calculated with five parameters (blue).

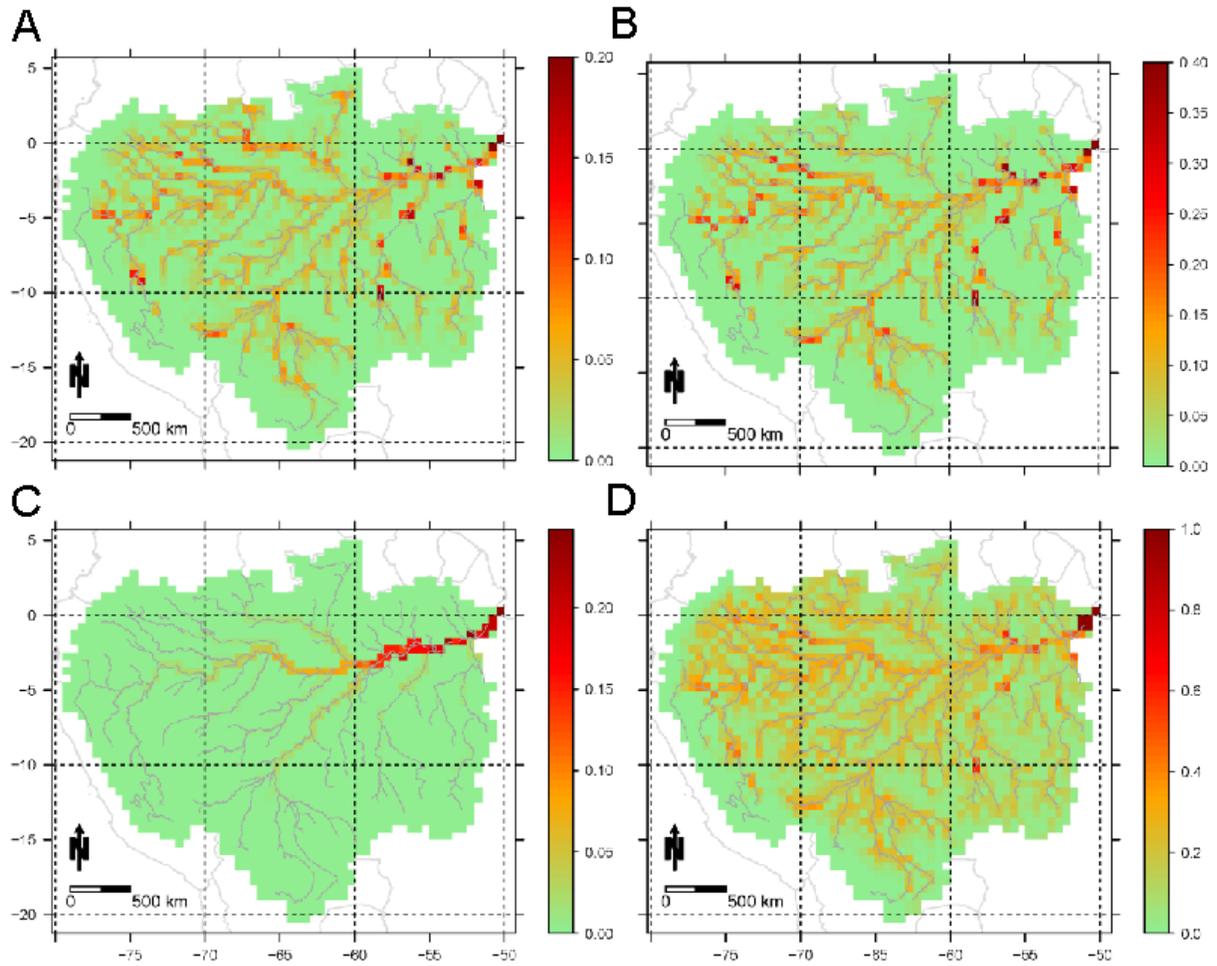
Table S3: Results of the redundancy analysis. Results for the first three RDA axes. Original value per axis (*axis*) and values proportional to the explained variability of the whole dataset (*prop*) with a general scaling constant of species scores of  $C_s = 3.9523$ .

	<i>RDA1</i>		<i>RDA2</i>		<i>RDA3</i>	
<b>Proportion explained</b>						
	0.540		0.189		0.063	
<b>Species scores (response variables)</b>						
	<i>axis</i>	<i>prop</i>	<i>axis</i>	<i>prop</i>	<i>axis</i>	<i>prop</i>
TOC concentration	-1.668	-0.228	0.4523	0.0216	-0.1601	-0.0026
POC concentration	-1.325	-0.181	0.3446	0.0165	0.8283	0.0132
DOC concentration	-1.564	-0.214	0.4296	0.0205	-0.5249	-0.0084
outgassed CO <sub>2</sub>	-1.204	-0.164	-1.5634	-0.0748	-0.0080	-0.0001
<b>Variable scores (explaining variables)</b>						
	<i>axis</i>	<i>prop</i>	<i>axis</i>	<i>prop</i>	<i>axis</i>	<i>prop</i>
<i>mobil<sub>litc</sub></i>	-0.5900	-0.3185	-0.77757	-0.14696	-0.00312	-0.00020
<i>mobil<sub>soilc</sub></i>	-0.1262	-0.0681	-0.17715	-0.03348	-0.00110	-0.00007
<i>mobil<sub>p</sub></i>	-0.0650	-0.0351	0.04598	0.00869	0.80759	0.05104
<i>decomp</i>	0.0452	0.0244	-0.03163	-0.00598	-0.58934	-0.03725
<i>respi</i>	0.7941	0.4287	-0.60111	-0.11361	0.08909	0.00563

#### 91 **4 Calibration and validation**

92 As a result of the sensitivity analysis, we calibrated values of the most important explaining  
93 variables (parameters)  $mobil_{litc}$ ,  $mobil_{soilc}$  and  $respi$  (Tables 3, 4 and S4). After calibration the  
94 Willmott's Index of Agreement, with 1 indicating complete agreement between observation and  
95 simulation results (Willmott, 1982), is 0.615, compared to 0.427 with the initial parameter values  
96 (Table 4). The calibrated rate of respiration ( $respi$ ) lies within the observed range, while the two  
97 other calibrated parameters ( $mobil_{litc}$ ,  $mobil_{soilc}$ ) are larger than observed values by a factor of 1.4  
98 and 5, respectively (Table 3). However, the observations available were only conducted in a  
99 *Várzea* ecosystem and  $mobil_{soilc}$  and are only estimated.

100 Spatial pattern and distribution of the carbon pools as calculated with the standard parameter  
101 setting are shown in Figure 4. The two organic carbon pools POC and DOC show the same  
102 spatial pattern with high amounts concentrated along the river, and only differ in the actual  
103 values with POC displaying half the amount of DOC (max.  $0.2 \times 10^8 \text{ g km}^{-2}$  vs. max.  
104  $0.4 \times 10^8 \text{ g km}^{-2}$ , Fig. S3). In contrast, the two inorganic carbon pools differ in their spatial  
105 pattern. The amount of inorganic carbon per cell (IC) increased up to  $0.25 \times 10^8 \text{ g km}^{-2}$  with  
106 increasing river discharge. The outgassed carbon is more homogeneously distributed in the  
107 catchment. Here, also the river network in combination with the floodplain can be identified.  
108 Therefore, the pattern is less pronounced than in the other carbon pools.

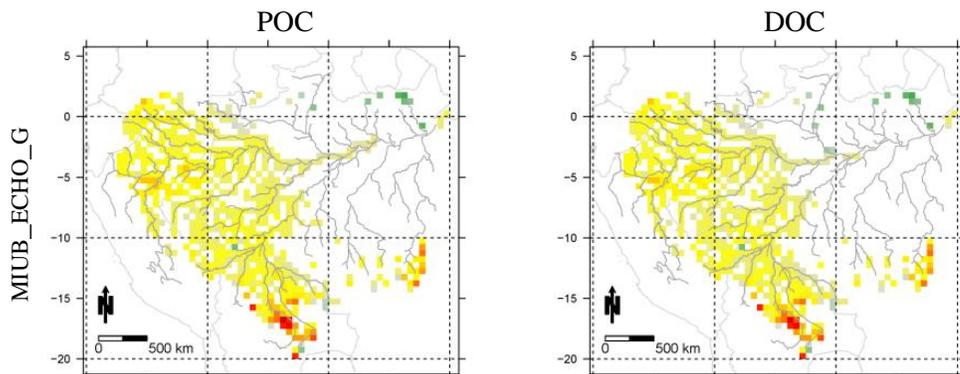


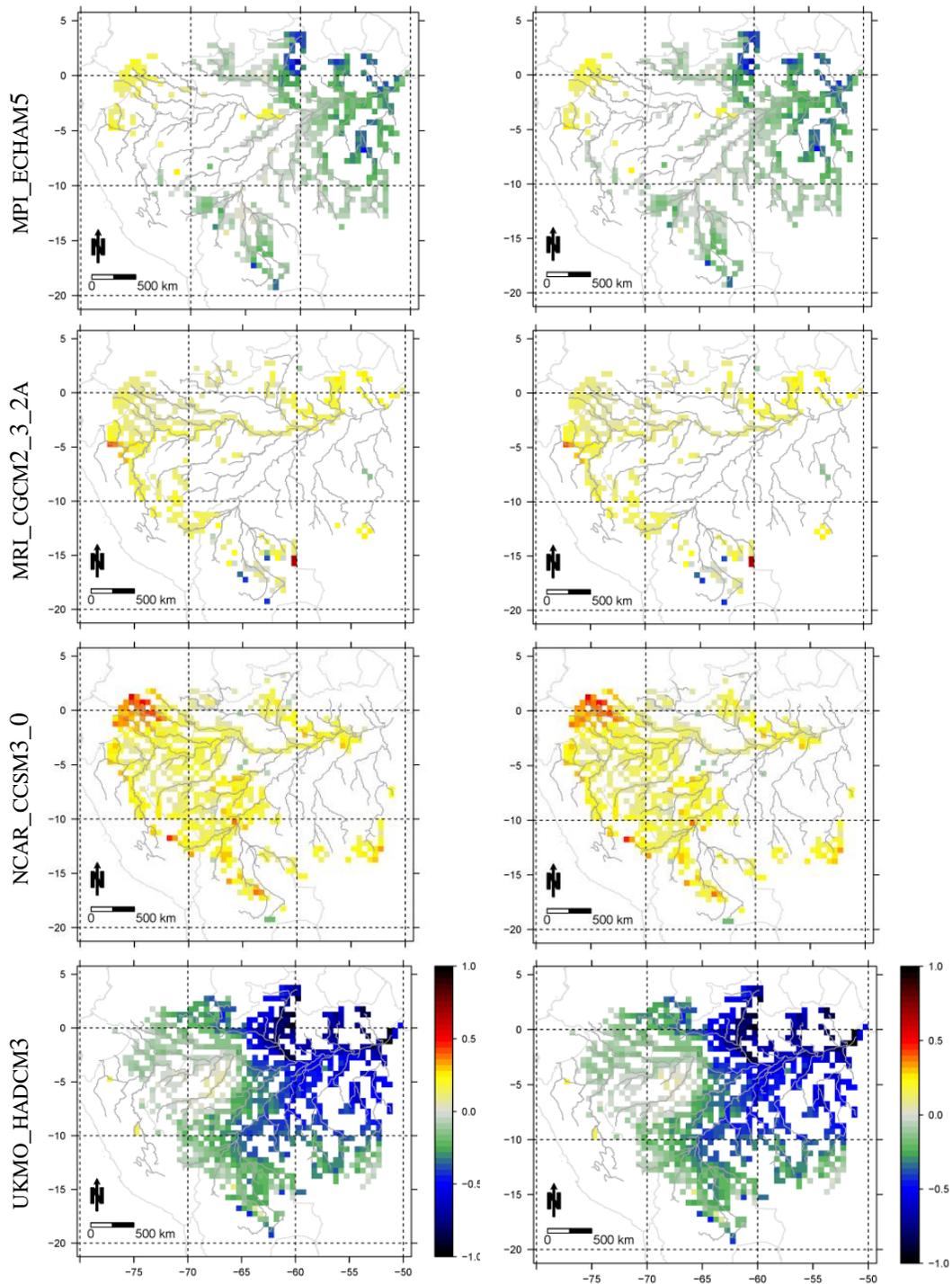
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Figure S3: Spatial distribution of the four carbon species used in the further analysis. Riverine particulate organic (POC), dissolved organic (DOC) and inorganic carbon (IC), and outgassed carbon [ $108 \text{ g km}^{-2}$ ] in 2003 obtained from simulations forced by CRU TS2.1.

116 **5 Additional maps showing similar patterns for POC and DOC**

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118 Figure S4: Changes in particulate organic carbon (POC) and dissolved organic carbon (DOC) caused by  
 119 climate change. Quotient ( $\log_{10}$ ) of mean future and mean reference carbon amount for each climate  
 120 model/scenario under emission scenario A1B. Positive values indicate an increase and negative values  
 121 indicate decrease.  
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123 **6 References**

- 124 Belger, L., Forsberg, B. R. and Melack, J. M.: Carbon dioxide and methane emissions from  
125 interfluvial wetlands in the upper Negro River basin, Brazil, *Biogeochemistry*, 105, 171–183,  
126 doi:10.1007/s10533-010-9536-0, 2011.
- 127 Cole, J. J., Pace, M. L., Carpenter, S. R. and Kitchell, J. F.: Persistence of net heterotrophy in  
128 lakes during nutrient addition and food web manipulations, *Limnol. Oceanogr.*, 45(8), 1718–  
129 1730, 2000.
- 130 Devol, A. H., Quay, P. D., Richey, J. E. and Martinelli, L. A.: The role of gas-rxchange in the  
131 inorganic carbon, oxygen, and Rn-222 budgets of the Amazon river, *Limnology and*  
132 *Oceanography*, 32(1), 235–248, 1987.
- 133 Furch, K. and Junk, W. J.: The chemical composition, food value, and decomposition of  
134 herbaceous plants, leaves, and leaf litter of floodplain forests, in *The Central Amazon*  
135 *Floodplain*, edited by W. J. Junk, pp. 187–205, Springer, Berlin, Germany., 1997.
- 136 Irmiler, U.: Litterfall and nitrogen turnover in an Amazonian blackwater inundation forest, *Plant*  
137 *and Soil*, 67(1-3), 355–358, 1982.
- 138 Johnson, M. S., Lehmann, J., Selva, E. C., Abdo, M., Riha, S. and Couto, E. G.: Organic carbon  
139 fluxes within and streamwater exports from headwater catchments in the southern Amazon,  
140 *Hydrological Processes*, 20(12), 2599–2614, 2006.
- 141 Junk, W. J. and Wantzen, K. M.: The flood pulse concept: New aspects, approaches and  
142 applications - An update, in *Proceedings of the Second International Symposium on the*  
143 *Management of large Rivers for Fisheries*, edited by R. L. Welcomme and T. Petr, pp. 117–140.,  
144 2004.
- 145 Lewin-Koh, N. J. and Bivand, R.: maptools: Tools for reading and handling spatial objects. R  
146 package version 0.8-7., 2011.
- 147 McClain, M. E. and Elsenbeer, H.: Terrestrial inputs to Amazon streams and internal  
148 biogeochemical processing, in *The Biogeochemistry of the Amazon Basin*, edited by M. E.  
149 McClain, R. L. Victoria, and J. E. Richey, pp. 185–208, Oxford University Press, New York.,  
150 2001.
- 151 Melack, J. M. and Forsberg, B.: Biogeochemistry of Amazon floodplain lakes and associated  
152 wetlands, in *The Biogeochemistry of the Amazon Basin and its Role in a Changing World*, pp.  
153 235–276, Oxford University Press, Eds. McClain, M. E.; Victoria, R. L.; Richey, J. E., 2001.
- 154 Neu, V., Neill, C. and Krusche, A. V.: Gaseous and fluvial carbon export from an Amazon forest  
155 watershed, *Biogeochemistry*, 105, 133–147, doi:10.1007/s10533-011-9581-3, 2011.
- 156 Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., O'Hara, R. B., Simpson, G. L., Solymos,  
157 P., Stevens, M. H. H. and Wagner, H.: vegan: Community Ecology Package. R package version  
158 1.17.11. [online] Available from: <http://CRAN.R-project.org/package=vegan>, 2011.

159 Poulter, B., Aragão, L., Heyder, U., Gumpenberger, M., Heinke, J., Langerwisch, F., Rammig,  
160 A., Thonicke, K. and Cramer, W.: Net biome production of the Amazon Basin in the 21st  
161 century, *Global Change Biology*, 16(7), 2062–2075, doi:10.1111/j.1365-2486.2009.02064.x,  
162 2009.

163 Rammig, A., Jupp, T., Thonicke, K., Tietjen, B., Heinke, J., Ostberg, S., Lucht, W., Cramer, W.  
164 and Cox, P.: Estimating the risk of Amazonian forest dieback, *New Phytologist*, 187(3), 694–  
165 706, doi:10.1111/j.1469-8137.2010.03318.x, 2010.

166 R Development Core Team and contributors worldwide, N. J.: stats: The R Stats Package version  
167 2.13.0., 2011.

168 Richey, J. E., Melack, J. M., Aufdenkampe, A. K., Ballester, V. M. and Hess, L. L.: Outgassing  
169 from Amazonian rivers and wetlands as a large tropical source of atmospheric CO<sub>2</sub>, *Nature*,  
170 416(6881), 617–620, doi:10.1038/416617a, 2002.

171 Silva, T. S. F., Costa, M. P. F. and Melack, J. M.: Annual net primary production of macrophytes  
172 in the Eastern Amazon floodplain, *Wetlands*, 29(2), 747–758, 2009.

173 Sippel, S. J., Hamilton, S. K. and Melack, J. M.: Inundation area and morphometry of lakes on  
174 the Amazon River floodplain, Brazil, *Archiv Fuer Hydrobiologie*, 123(4), 385–400, 1992.

175 Willmott, C. J.: Some comments on the evaluation of model performance, *Bulletin American*  
176 *Meteorological Society*, 63(11), 1309–1313, 1982.

177 Worbes, M.: The forest ecosystem of the floodplains, in *The Central Amazon Floodplain*, edited  
178 by W. J. Junk, pp. 223–265, Springer, Berlin, Germany., 1997.

179